Some Studies on the Uses of 2-Bromoethyl and 2-Iodoethyl Ester Blocking Groups in **Peptide Synthesis: Samarium Diiodide-Mediated Deprotection**

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Introduction

In connection with studies, currently underway in our laboratory, that are directed toward the synthesis of compounds related to the glycopeptide antibiotics ristocetin and vancomycin,¹ we required a carboxylate blocking group that is sufficiently robust to withstand a number of synthetic operations elsewhere in the molecule, yet can be removed under mild neutral conditions. The latter restriction is due to the fact that the target molecules contain a number of arylglycine residues that are prone to racemization under mildly basic conditions, and that tert-butoxycarbonyl and/or benzyloxycarbonyl groups were projected for amine protection during their assembly. A potentially useful candidate for our purposes is the 2-bromoethyl ester, which has been reported to be removable under a variety of nonbasic reaction conditions.² We were soon to learn, however, that this deprotection is highly problematic. In almost every case that we have studied, the 2-bromoethyl ester was converted to a 2-hydroxyethyl ester, probably via a classical neighboring group participation involving the carboxyl oxygen, which facilitates hydrolysis of the bromide.³ It has been reported that the 2-bromoethyl ester can be converted, via Finkelstein reaction, to a 2-iodoethyl ester.^{2c} The latter is somewhat easier to cleave reductively (Zn powder) and both twostep and one-pot procedures have been described for unmasking the carboxylate group.^{2c,4} In our hands, this tactic has also led to formation of 2-hydroxyethyl esters, either exclusively or as the major product.³ Magnus and co-workers have described similar difficulties during the deprotection of 2-chloroethyl carbamates, a problem that was solved by using samarium diiodide as the reducing agent.⁵ This paper reports on the use of SmI_2 for the deprotection of 2-iodoethyl esters under anhydrous conditions, which are compatible with the presence of arylglycines as well as a number of amine blocking groups, leading to the carboxylic acids in reproducibly excellent yields.

Results and Discussion

A number of 2-bromoethyl esters were prepared from the corresponding acids by following the standard method

(1) Pearson, A. J.; Park, J. G. J. Org. Chem. 1992, 57, 1744.

using 2-bromoethanol/(DCC, pyridine) (Table 1). Direct deprotection of these esters by using SmI₂ was found to be extremely slow and inefficient, and so they were converted to 2-iodoethyl esters via Finkelstein reaction using NaI (5.0 equiv) in dry acetone for 7 h under nitrogen. This halide conversion was easily confirmed by ¹H and ¹³C NMR. Several 2-iodoethyl esters were also prepared directly from reaction of the carboxylic acid with 2-iodoethanol in the presence of DCC.

In the initial studies of deprotection by using SmI_2 , N-Boc-p-chloro-D-phenylalanine 2-iodoethyl ester (1b) was used to determine the optimum reaction conditions. The reasons for choosing 1b as model compound are that it has a relatively sensitive protecting group (N-Boc) and it is a chiral amino acid which is very closely related to intermediates that are currently being employed in our studies on vancomycin and ristocetin A. After reaction of 1b with SmI_2 (7.0 equiv) at rt under deoxygenated, dry argon for 1 day, 75% was deprotected (¹H NMR); prolonged stirring (2 days) with excess of SmI_2 (10.0 equiv) did not lead to complete deprotection. At slightly higher temperature $(35 \,^{\circ}\text{C}, 7.0 \,\text{equiv of } \text{SmI}_2)$ the reaction was determined to be complete after 10 h from the ¹H NMR of crude product; at 40 °C, several products were observed, presumably due to instability of the N-Boc protecting group. For a comparison of N-Cbz and N-Boc, we investigated the deprotection of 2, which showed that N-Cbz is stable toward SmI₂ at 40 °C. The optimized reaction conditions



For all structures: (a) $R = CH_2CH_2Br$; (b) $R = CH_2CH_2I$; (c) R = H

thus obtained (A: SmI₂ (6.0 equiv), 40 °C, 8 h; B: SmI₂ (7.0 equiv), 35 °C, 10 h) were used with several different

^{(2) (}a) Greene, T. W. Protective Groups in Organic Synthesis; John Wiley: New York, 1981. (b) Anderson, L. C; Pinnick, H. W. J. Org. Chem. , 43, 3417. (c) Kunz, H.; Buchholz, M. Chem. Ber. 1979, 112, 2145. (d) Ho, T. L. Synthesis 1975, 510. (e) Ho, T. L. Synthesis 1974, 715. For related methods, see Ho, T. L. Synth. Commun. 1978, 8, 301. (f) Scheffold, R.; Amble, E. Angew. Chem., Int. Ed. Engl. 1980, 19, 629. (g) Jacobson, R. M.; Clader, J. W. Synth. Commun. 1979, 9, 57. (h) Fried, J.; Sabo, T. F. J. Am. Chem. Soc. 1957, 79, 1130. (i) Joaquina, M; Trigo, S. A. A.; Santon, M. I. A. O. Collect. Czech. Chem. Commun. 1988, 53, 2787. (j) (3) Park, J. G. Ph.D. Thesis, Case Western Reserve University, 1991.

⁽⁴⁾ Kunz, H.; Buchholz, M. Liebigs Ann. Chem. 1983, 11, 1859.

⁽⁵⁾ Magnus, P.; Ananthamarayan, T. P.; Gallagher, T. J. Chem. Soc., Chem. Commun. 1982, 709.

 Table 1. Yields for the Preparation and Deprotection of 2-Iodoethyl Esters*

entry	substrate	yield of b (%)	yield for $\mathbf{b} \rightarrow \mathbf{c}$ (%)
1	1 ^b	75	82
2	2	87	88
3	3	86	93
4	4	91	91
5	5	89	94
6	6	86	89
7	7	83	76
8	8°	95	70
9.	9d	82	38
10	10 ^b	80	90
11	11 ^b	75	93
12	12 ^b	73	88

^a Methods and spectroscopic data are given in the Experimental Section; for all structures: (a) $R = CH_2CH_2Br$; (b) $R = CH_2CH_2I$; (c) R = H. ^b These iodoethyl esters were prepared directly from the corresponding acid (RCO₂H/iodoethanol/DCC), as described for compound 1b. ^c The preparation of this compound is described in ref 8. ^d The preparations of 7a and 9a are described in refs 1 and 3.

compounds, and the results are summarized in Table 1. For simple 2-iodoethyl ester derivatives, reaction conditions A worked well and gave products of high purity in excellent yields. For amino acid derivatives, reaction conditions A (for N-Cbz) or B (for N-Boc) gave good to excellent yields. We next turned our attention to the possibility of racemization during the deprotection reaction. For this investigation, the racemization-prone phenylglycine derivative 2 was chosen as a model compound. Conversion of 2c ($[\alpha]^{22}_{D} = -121.2^{\circ}$, c 0.58, EtOAc) to 2a (2-bromoethanol, DCC, HOBT, DMF, CH₂Cl₂, 0 °C, 17 h, 86%) then to 2b (NaI, acetone, Δ), followed by deprotection of 2b, returned 2c without change in optical rotation ($[\alpha]^{22}$ _D $= -121.0^{\circ}$, c 0.48, EtOAc), showing that no racemization occurs during the entire protection/halide exchange/ deprotection sequence.

Conclusions

The 2-iodoethyl ester protecting group is removed successfully with SmI_2 under mild conditions. Theoretically, 2.0 equiv of SmI_2 is sufficient for this deprotection reaction, and the requirement for a large excess of this reagent may be due to its instability. In this context it may be noted that the deprotection does not proceed to completion unless oxygen is rigorously excluded, owing to competing oxidation of the samarium reagent. A twostep sequence (Finkelstein reaction, followed by deprotection with SmI_2) provides useful methodology for the deprotection of 2-bromoethyl esters. The latter protecting group is more stable than iodoethyl toward various reaction conditions, but its direct removal is problematic.

Experimental Section

 SmI_2 (0.1 M in THF) was purchased from Aldrich Co. and used directly. All reactions using SmI_2 were performed under dry oxygen-free argon. All spectroscopic and general procedures were as described previously.¹

General Procedure for the Preparation of 2-Bromoethyl Esters and Direct Preparation of 2-Iodoethyl esters. See the preparation of 6a and 1b, respectively.

General Procedure for Finkelstein Reaction. See the preparation of 9b.

General Procedure for the Deprotection of Iodoethyl Esters Using SmI₂. See the preparation of 2c and 7c.

N-[(1,1-Dimethylethoxy)carbonyl]-4-chloro-D-phenylalanine 2-Iodoethyl Ester (1b). To a precooled (0 °C), stirred solution of N-Boc-4-chloro-D-phenylalanine (556 mg, 1.86 mmol) in 20 mL of CH_2Cl_2 under N_2 were added 174 μL (1.2 equiv) of 2-iodoethanol followed by 301 μ L of pyridine (2.0 equiv) by syringe. The mixture was stirred for 10 min at 0 °C and then 422 mg of DCC (1.1 equiv) was added in one portion. The resulting mixture was stirred for 16 h at 0 °C under N₂ in the dark. The reaction was quenced by the addition of 25 mg (0.15 equiv) of oxalic acid in 0.5 mL of THF and then it was allowed to come to rt and stirred for 30 min. The solid was filtered off and the filter cake was washed well with CH_2Cl_2 (10 mL \times 3). The combined organic extracts were washed with 0.1 N HCl and brine and dried over MgSO₄, and the solvent was removed in vacuo to give a solid residue. Purification by flash chromatography on silica gel (Hex/EtOAc, 7/3) gave 629 mg (75%) of 1b as a white solid: mp 80.5-81.5 °C; Rf 0.37 (Hex/EtOAc, 7/3); IR (CHCl₃) 3438, 3026, 2982, 2934, 1744, 1711, 1493 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 7.28 (d, 2 H, J = 8.3 Hz), 7.11 (d, 2 H, J = 8.3 Hz), 4.96 (d, 1 H, J = 7.2 Hz), 4.59-4.57 (m, 1 H), 4.39-4.34 (m, 2 H), 3.28-3.00 (m, 4 H, CO₂CH₂CH₂I overlapping with ArCHHCH, ArCHHCH), 1.42 (8, 9 H); ¹³C NMR (75 MHz, CDCl₂) & 171.1. 155.0, 134.4, 133.0, 130.7, 128.7, 80.2, 65.4, 54.2, 37.7, 28.3, -0.6; HRMS calcd for C₁₆H₂₁NO₂ClI 453.0206, found 453.0213.

N-[(1,1-Dimethylethoxy)carbonyl]-4-chloro-D-phenylalanine (1c): yield 82%; mp 106.5–108.5 °C; $[\alpha]^{22}_D - 28.7^{\circ}$ (c 0.2, EtOAc); R_f 0.35 (EtOAc/MeOH, 9/1); IR (CHCl₃) 3438, 3026, 2982, 1719, 1653, 1493, 1221 cm⁻¹; ¹H NMR (300 MHz, DMSO d_6) δ 12.62 (bs, 1 H), 7.32–7.24 (m, 4 H), 7.10 (d, 1 H, J = 8.8 Hz), 4.11–4.04 (m, 1 H), 3.00 (dd, 1 H, J = 13.7, 3.8 Hz), 2.80 (dd, 1 H, J = 13.7, 10.9 Hz); ¹³C NMR (75 MHz, DMSO- d_6) δ 173.4, 155.4, 137.1, 131.0, 128.0, 78.1, 55.0, 35.8, 28.1.

N-[(Phenylmethoxy)carbonyl]-D-phenylglycine 2-bromoethyl ester (2a): yield 86%; mp 62–64 °C; $[\alpha]^{22}_D$ -50.1° (c 0.74, EtOAc); R_f 0.30 (Hex/EtOAc, 7/3); IR (CHCl₃) 3435, 3036, 2946, 1723, 1500, 1216 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.36–7.25 (m, 10 H), 5.78 (d, 1 H, J = 7.3 Hz, PhCH₂OCONHCH), 5.41 (d, 1 H, J = 7.3 Hz, PhCH₂OCONHCH), 5.13 (d, 1 H, J = 12.2 Hz, PhCHHOCO), 5.08 (d, 1 H, J = 12.2 Hz, PhCHHOCO), 4.40 (m, 2 H, CO₂CH₂CH₂Br), 3.41 (m, 2 H, CO₂CH₂CH₂Br); ¹³C NMR (75 MHz, CDCl₃) δ 170.4, 155.3, 136.0, 129.0, 128.7, 128.5, 128.2, 127.2, 67.2, 64.8, 58.0, 27.7; HRMS calcd for C₁₈H₁₈NO₄Br 391.0420, found 391.0408.

N-[(Phenylmethoxy)carbonyl]-D-phenylglycine 2-iodoethyl ester (2b): yield 87%; mp 68.5–70.5 °C; $[\alpha]^{25}_{D}$ –55.3° (c 0.6, CHCl₃); R_f 0.36 (Hex/EtOAc, 7/3); IR (CHCl₃) 3435, 3014, 2967, 1742, 1723, 1602, 1517 cm⁻¹; ¹H NMR (300 MHz, CDCl₃); δ 7.37–7.26 (m, 10 H), 5.78 (d, 1 H, J = 7.1 Hz), 5.39 (d, 1 H, J = 7.1 Hz), 5.13 (d, 1 H, J = 12.1 Hz), 5.08 (d, 1 H, J = 12.1 Hz), 4.40–4.33 (m, 2 H), 3.49–3.17 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) δ 170.2, 155.3, 136.1, 136.0, 129.0, 128.7, 128.5, 128.2, 127.2, 67.2, δ 55.6, 58.0, -0.9; HRMS calcd for C₁₈H₁₈NO₄I 439.0282, found 439.0283.

N-[(Phenylmethoxy)carbonyl]-D-phenylglycine (2c). To a stirred, precooled (0 °C) solution of 2-iodoethyl ester (2b, 10 mg. 0.18 mmol) in 10 mL of THF was added 10.9 mL (6.0 equiv, 0.1 M in THF) of SmI_2 by syringe. The resulting mixture was warmed to 40 °C and stirred for 8 h under deoxygenated argon. The mixture was cooled to rt, diluted with 30 mL of EtOAc, and quenched by the addition of 10 mL of 0.1 N HCl. The organic layer was separated and the aqueous layer was washed with EtOAc $(10 \text{ mL} \times 2)$. The combined organic extracts were washed with 10 mL of 0.2 M sodium thiosulfate and brine and dried over MgSO₄, and the solvent was removed in vacuo to give a yellow residue. Purification by short column chromatography on silica gel (1 \times 15 cm, EtOAc/MeOH, 95/5) afforded 46 mg (88%) of 2c as a white solid: mp 127.5–129.5 °C; $[\alpha]^{24}_{D}$ –121.0° (c 0.48, EtOAc); R_{f} 0.21 (EtOAc/MeOH, 9/1); IR (CHCl₃) 3436, 3026, 1725, 1667, 1497 cm⁻¹; ¹H NMR (300 MHz, DMSO-d₆) δ 12.86 (bs, 1 H, PhCHCO₂H), 8.11 (d, 1 H, J = 8.1 Hz, PhCH₂OCONH), 7.41-7.30 (m, 10 H), 5.16 (d, 1 H, J = 8.1 Hz, PhCHCO₂H), 5.04 (s, 2 H, PhCH₂OCONH); ¹³C NMR (75 MHz, DMSO-d₆) δ 172.0, $155.8, 137.1, 136.9, 128.4, 127.7, 65.6, 58.0; HRMS calcd for <math display="inline">\rm C_{16}H_{15}\text{-}$ NO₄ 285.1001, found 285.1001

3-(4-Methoxyphenyl)propionic acid 2-bromoethyl ester (3a): yield 72%; R_f 0.56 (Hex/EtOAc, 6/4); IR (CHCl₃) 3014, 1734, 1514 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.11 (d, 2 H, J =8.6 Hz), 6.82 (d, 2 H, J = 8.6 Hz), 4.35 (t, 2 H, J = 6.1 Hz, CO₂CH₂-CH₂Br), 3.76 (s, 3 H, OCH₃), 3.45 (t, 2 H, J = 6.1 Hz, OCH₂CH₂- Br), 2.90 (t, 2 H, J = 7.7 Hz, ArCH₂CH₂CO₂), 2.63 (t, 2 H, J = 7.7 Hz, ArCH₂CH₂CO₂); ¹³C NMR (75 MHz, CDCl₃) δ 172.3, 158.0, 132.1, 129.1, 113.8, 63.6, 55.1, 35.8, 29.9, 28.6; HRMS calcd for C₁₂H₁₆O₃Br 286.0205, found 286.0203.

3-(4-Methoxyphenyl) propionic acid 2-iodoethyl ester (3b): yield 86%; R_f 0.49 (Hex/EtOAc, 7/3); IR (CHCl₃) 3020, 1734, 1514 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.12 (d, J = 8.6 Hz, 2 H), 6.83 (d, J = 8.6 Hz, 2 H), 4.32 (t, J = 6.9 Hz, 2 H, OCH₂CH₂I), 3.78 (s, 3 H), 3.25 (t, J = 6.9 Hz, 2 H, OCH₂CH₂I), 2.91 (t, J =7.7 Hz, 2 H, ArCH₂CH₂CO₂), 2.63 (t, J = 7.7 Hz, 2 H, ArCH₂CH₂CO₂); ¹³NMR (75 MHz, CDCl₃) δ 172.3, 158.1, 132.3, 129.2, 113.9, 64.5, 55.3, 36.0, 30.0, 0.3; HRMS calcd for C₁₂H₁₆O₃I 334.0068, found 334.0067.

3-(4-Methoxyphenyl)propionic acid (3c): yield 93%; mp 102.5-104.0 °C (Aldrich catalogue: 98-100 °C); R_f 0.56 (Hex/ EtOAc, 2/8); IR (CHCl₃) 3011, 2957, 1711, 1612, 1514, 1248 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.98 (bs, 1 H), 7.12 (d, 2 H, J =8.0 Hz), 6.83 (d, 2 H, J = 8.0 Hz), 3.79 (s, 3 H), 2.90 (t, 2 H, J =7.7 Hz), 2.65 (t, 2 H, J = 7.7 Hz).

3-[3-(Benzyloxy)-4-methoxyphenyl]propionic acid 2-bromoethyl ester (4a): yield 83%; mp 44.0–45.5 °C; R_f 0.35 (Hex/ EtOAc, 8/2); IR (CHCl₃) 3016, 1737, 1515 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.27 (m, 5 H), 6.84–6.74 (m, 3 H), 5.13 (s, 2 H, OCH₂Ph), 4.34 (t, 2 H, J = 6.2 Hz), 3.86 (s, 3 H), 3.45 (t, 2 H, J = 6.2 Hz), 2.86 (t, 2 H, J = 7.7 Hz), 2.60 (t, 2 H, J = 7.7 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 172.4, 148.3, 148.1, 137.1, 132.8, 128.5, 127.8, 127.3, 120.8, 114.5, 111.9, 71.0, 63.7, 56.1, 35.8, 30.3, 28.7; HRMS calcd for C₁₉H₂₁O₄Br 392.0624, found 392.0629.

3-[3-(Benzyloxy)-4-methoxyphenyl]propionic acid 2-iodoethyl ester (4b): yield 91%; mp 52.5–54.0 °C; R_f 0.55 (Hex/ EtOAc, 8/2); IR (CHCl₃) 3016, 1737, 1515 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.26 (m, 5 H), 6.84–6.74 (m, 3 H), 5.13 (s, 2 H), 4.29 (t, 2 H, J = 6.8 Hz), 3.86 (s, 3 H) 3.23 (t, 2 H, J = 6.8 Hz), 2.86 (t, 2 H, J = 7.7 Hz), 2.59 (t, 2 H, J = 7.7 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 148.3, 148.1, 137.1, 132.8, 128.5, 127.8, 127.3, 120.8, 114.5, 111.9, 71.0, 64.4, 56.1, 35.9, 30.4, 0.4; HRMS calcd for C₁₉H₂₁O₄I 440.0486, found 440.0487.

3-[3-(Benzyloxy)-4-methoxyphenyl]propionic acid (4c): yield 91%; mp 122.5–124.5 °C; R_f 0.41 (Hex/EtOAc, 2/8); IR (CHCl₃) 3522, 3017, 2935, 1710, 1591, 1515, 1256, 1211 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.26 (m, 5 H), 6.84–6.74 (m, 3 H), 5.13 (s, 2 H), 3.86 (s, 3 H), 2.85 (t, 2 H, J = 7.7 Hz), 2.60 (t, 2 H, J = 7.7 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 179.0, 148.2, 148.0, 137.1, 132.6, 128.5, 127.8, 127.3, 120.7, 114.5, 111.9, 71.0, 56.0, 35.7, 30.0; HRMS calcd for C₁₇H₁₈O₄ 286.1205, found 286.1210.

3-(Benzyloxy)-4-methoxyphenylacetic acid 2-bromoethyl ester (5a): yield 71%; mp 54.0–55.5 °C; R_f 0.32 (Hex/EtOAc, 8/2); IR (CHCl₃) 3019, 2944, 1736, 1515 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.25 (m, 5 H), 6.86–6.82 (m, 3 H), 5.15 (s, 2 H), 4.35 (t, 2 H), 3.87 (s, 3 H), 3.54 (s, 2 H), 3.44 (t, 2 H, J = 6.2 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 171.2, 148.9, 148.1, 137.0, 128.5, 127.8, 127.3, 125.9, 122.0, 115.0, 111.8, 70.9, 64.0, 56.0, 40.6, 28.6; HRMS calcd for C₁₈H₁₉O₄Br 378.0467, found 378.0464.

3-(Benzyloxy)-4-methoxyphenylacetic acid 2-iodoethyl ester (5b): yield 89%; mp 59.5–61.5 °C; R_f 0.35 (Hex/EtOAc, 7/3); IR (CHCl₃) 3019, 2936, 1734, 1515 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.25 (m, 5 H), 6.86–6.84 (m, 3 H), 5.16 (s, 2 H), 4.28 (t, 2 H, J = 6.8 Hz), 3.86 (s, 2 H), 3.53 (s, 2 H), 3.20 (t, 2 H, J= 6.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 171.0, 148.9, 148.1, 137.0, 128.5, 127.8, 127.2, 125.9, 122.0, 115.0, 111.8, 70.9, 64.7, 56.0, 40.6, 0.2; HRMS calcd for C₁₈H₁₉O₄I 426.0330, found 426.0319.

3-(Benzyloxy)-4-methoxyphenylacetic acid (5c): yield 94%; mp 122.0–123.5 °C (lit.⁶ 123.0–125.0 °C); R_f 0.47 (CHCl₃/MeOH, 10/1); IR (CHCl₃) 3020, 2936, 1711, 1515, 1443 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.25 (m, 5 H), 6.85 (s, 3 H), 5.13 (s, 2 H), 3.87 (s, 3 H), 3.55 (s, 2 H).

N-[(Phenylmethoxy)carbonyl]glycine 2-bromoethyl ester (6a). To a stirred, precooled (0 °C) solution of N-Cbz-glycine (1.00 g, 4.78 mmol) in 40 mL of dry CH₂Cl₂ were added 407 μ L (1.2 equiv) of 2-bromoethanol, followed by 773 μ L (2.0 equiv) of pyridine by syringe. The mixture was stirred for 10 min at 0 °C, 1.09 g (1.1 equiv) of DCC was added in one portion, and the resulting mixture was stirred for 16 h at 0 °C under N₂. The reaction was quenched with 143 μ L (0.15 equiv) of 5 M oxalic acid in THF, allowed to come to rt, and stirred further for 30 min. The mixture was filtered and the filter cake was washed well with CH_2Cl_2 (10 mL × 3). The combined organic extracts were washed with dilute HCl (0.1 N, 20 mL) and brine and dried over MgSO₄. After removal of solvent in vacuo, the crude product was flash-chromatographed (silica gel, Hex/EtOAc, 7/3) to give 1.27 g (84%) of pure product 6a as a pale yellow solid: mp 59.0-60.0 °C; R_f 0.21 (Hex/EtOAc, 7/3); IR (CHCl₃) 3452, 3020, 1752, 1724, 1517, 1220 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.36-7.29 (m, 5 H), 5.26 (bs, 1 H, PhCH₂OCONH), 5.14 (s, 2 H, PhCH₂-OCONH), 4.46 (t, 2 H, J = 6.1 Hz, CO₂CH₂CH₂Br), 4.04 (d, 2 H, J = 5.6 Hz, NHCH₂CO₂), 3.51 (t, 2 H, J = 6.1 Hz, CO₂CH₂CH₂-Br); ¹³C NMR (75 MHz, CDCl₃) δ 169.6, 156.2, 136.1, 128.5, 128.2, 128.1, 67.1, 64.5, 42.6, 28.1; HRMS calcd for C₁₂H₁₄NO₄Br 315.0107, found 315.0104.

N-[(Phenylmethoxy)carbonyl]glycine 2-iodoethyl ester (**6b**): yield 86%; mp 85.5–87.0 °C; R_f 0.26 (Hex/EtOAc, 7/3); IR (CHCl₃) 3447, 3021, 1751, 1724, 1517 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.36–7.26 (m, 5 H), 5.24 (bs, 1 H, PhCH₂OCONH), 5.14 (s, 2 H, PhCH₂OCO), 4.41 (t, 2 H, J = 6.8 Hz, CO₂CH₂CH₂I), 4.02 (d, 2 H, J = 5.7 Hz, NHCH₂CO₂), 3.29 (t, 2 H, J = 6.8 Hz, CO₂-CH₂CH₂I); ¹³C NMR (75 MHz, CDCl₃) δ 169.4, 156.2, 136.1, 128.5, 128.2, 128.1, 67.2, 65.3, 42.7, -0.5; HRMS calcd for C₁₂H₁₄NO₄I 362.9969, found 362.9967.

N-[(Phenylmethoxy)carbony]glycine (6c): yield 89%; mp 116.5–118.5 °C (lit.⁷ 118–119 °C); R_f 0.25 (EtOAc/MeOH, 9/1); IR (CHCl₃) 3451, 3018, 1726, 1517 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.37–7.24 (m, 5 H), 5.23 (bs, 1 H, OCONHCH₂), 5.14 (s, 2 H, ArCH₂OCO), 4.06 (d, 2 H, J = 6.7 Hz, NHCH₂CO₂H).

(*R*)-4-O-[3-[1-[[(Phenylmethoxy)carbonyl]amino]-1-(methoxycarbonyl]-D-tyrosine α -(2-Bromoethyl) Ester (7a). Obtained as an oily foam; $[\alpha]^{24}_D$ -9.2° (c 0.47, CHCl₃); R_f 0.50 (Hex/EtOAc, 6/4); IR (CHCl₃) 3434, 3020, 1742, 1718, 1506 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.33-6.88 (m, 13 H), 5.96 (d, 1 H, J = 7.2 Hz, PhCH₂OCONHCH), 5.36 (d, 1 H, J = 7.2 Hz, PhCH₂OCONHCH), 5.36 (d, 1 H, J = 7.2 Hz, PhCH₂OCONHCH), 5.36 (d, 1 H, J = 7.2 Hz, PhCH₂OCONHCH), 5.14-5.04 (m, 3 H, PhCH₂OCO overlapping with NHCO₂C (CH₃)₃), 4.61-4.57 (m, 1 H, ArCH₂CHCO₂), 4.42-4.36 (m, 2 H, CO₂CH₂-CH₂Br), 3.71 (s, 3 H, CO₂CH₃), 3.4-3.00 (m, 2 H, ArCH₂CHCO₂), 1.42 (s, 9 H, NHCO₂C-(CH₃)₃); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.9, 157.5, 155.7, 155.3, 158.4, 136.0, 131.0, 130.6, 130.1, 128.4, 128.1, 121.7, 119.0, 118.4, 117.5, 80.0, 67.1, 64.5, 57.5, 54.4, 52.8, 37.4, 28.2

(*R*)-4-[3-[1-[[(Phenylmethoxy)carbonyl]amino]-1-(methoxycarbonyl)methyl]phenoxy]-*N*-[(1,1-dimethylethoxy)carbonyl]-D-tyrosine α -(2-Iodomoethyl) Ester (7b): yield 83%; oily foam; $[\alpha]^{25}_{D}$ -5.5° (c 0.55, CHCl₃); R_{f} 0.51 (Hex/EtOAc, 6/4); IR (CHCl₃) 3436, 3019, 1743, 1718, 1506 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.34-6.89 (m, 13 H), 5.89 (d, 1 H, J = 7.4 Hz), 5.36 (d, 1 H, J = 7.4 Hz), 5.11-5.00 (m, 3 H), 4.63-4.57 (m, 1 H), 4.39-4.34 (m, 2 H), 3.78 (s, 3 H), 3.27-3.22 (m, 2 H), 3.17-3.01 (m, 2 H), 1.42 (s, 9 H); ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 170.9, 157.6, 155.8, 155.2, 155.0, 138.5, 136.0, 131.0, 130.7, 130.2, 128.5, 128.1, 121.8, 119.1, 118.5, 117.6, 80.1, 67.1, 65.3, 57.6, 54.4, 52.9, 37.6, 28.3, -0.4.

(R)-4-[3-[1-[[(Phenylmethoxy)carbonyl]amino]-1-(methoxycarbonyl)methyl]phenoxy]-N-[(1,1-dimethylethoxy)carbonyl]-D-tyrosine (7c). To a stirred, precooled (0 °C) solution of 7b (84 mg, 0.11 mmol) in 10 mL of freshly distilled THF was added 8.1 mL (7.0 equiv) of SmI_2 (0.1 M in THF) by syringe, and the resulting mixture was warmed to 35 °C for 10 h under deoxygenated argon. The reaction mixture was cooled to rt, diluted with 20 mL of EtOAc, and quenched by the addition of 10 mL of 0.1 N HCl. The organic layer was separated and the aqueous layer was washed well with EtOAc (10 mL \times 2). The combined organic extracts were washed with 10 mL of 0.2 M sodium thiosulfate solution, washed with brine, dried over MgSO4, and concentrated in vacuo to give a solid residue. Purification by short column chromatography on silica gel $(1 \times 15 \text{ cm}, \text{Hex}/$ EtOAc, 2/8) gave 50 mg (76%) of pure acid 7c: $[\alpha]^{26}_{Hg,365}$ -1.8° (c 0.5, CH₃CN); R_f 0.25 (EtOAc/MeOH, 9/1); IR (CHCl_s) 3433, 3028, 3012, 2982, 1750, 1716, 1590, 1506, 1223 cm⁻¹; ¹H NMR (300

⁽⁶⁾ Pearson, A. J.; Shin, H. Tetrahedron 1992, 48, 7529.

⁽⁷⁾ Sennyey, G.; Barcelo, G.; Senet, J. P. Tetrahedron Lett. 1986, 27, 5375.

MHz, $CD_3CN + CDCl_3$) δ 7.36–6.90 (m, 13 H), 6.39 (d, 1 H, J = 7.2 Hz), 5.40 (d, 1 H, J = 7.2 Hz), 5.28 (d, 1 H, J = 7.7 Hz), 5.07 (s, 2 H), 4.35 (bs, 1 H), 3.68 (s, 3 H), 3.13 (dd, 1 H, J = 14.5, 4.3 Hz), 2.91 (dd, 1 H, J = 14.5, 8.3 Hz).

(S)-3-[4-[2-[[(1,1-Dimethylethoxy)carbonyl]amino]-3-oxo-3-(2-iodoethoxy)propy]phenoxy]-O-methyl-N-[O-methyl- $N-[(phenylmethoxy)carbonyl]-L-tyrosyl]-L-tyrosine \alpha-Me$ thyl Ester (8b). Details for the preparation of this compound are given elesewhere:⁸ mp 63.5-65.5 °C; [α]²⁶_D +34.3° (c 0.58, CHCl₃); R_f 0.19 (Hex/EtOAc, 6/4); IR (CHCl₃) 3427, 3028, 2983, 1742, 1713, 1683, 1612, 1505, 1228 cm⁻¹; ¹H NMR (300 MHz, $CDCl_{s}$) δ 7.34–6.59 (m, 16 H), 6.27 (d, 1 H, J = 7.1 Hz), 5.33 (bs, 1 H), 5.12-5.04 (m, 3 H), 4.72-4.68 (m, 1 H), 4.56-4.54 (m, 1 H), 4.36-4.32 (m, 3 H), 3.77 (s, 3 H), 3.76 (s, 3 H), 3.60 (s, 3 H), 3.25-3.20 (m, 2 H), 3.11-2.85 (m, 6 H), 1.41 (s, 9 H); ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 171.1, 170.4, 158.6, 157.1, 155.8, 155.1, 150.6, 144.4, 136.1, 130.4, 130.3, 129.7, 128.5, 128.4, 128.1, 127.9, 125.7, 122.1, 116.9, 114.0, 112.8, 80.0, 67.0, 65.2, 56.1, 55.9, 55.1, 54.4, 53.3, 52.3, 37.5, 37.3, 37.0, 28.2, -0.4. Anal. Calcd for C45H52N3O12I: C, 56.65; H, 5.50. Found: C, 56.56; H, 5.43.

(S)-3-[4-[2-Carboxy-2-[[(1,1-dimethylethoxy)carbonyl]amino]ethyl]phenoxy]-O-methyl-N-[O-methyl-N-[(phenylmethoxy)carbonyl]-L-tyrosyl]-L-tyrosine α -Methyl Ester (8c): yield 70%; $[\alpha]^{23}_{Hg,365}$ +30.4° (c 0.38, MeOH, lit.⁹ +27.6° (c 0.54, MeOH)); R_f 0.17 (EtOAc/MeOH, 9/1); IR (CHCl₃) 3412, 3054, 2985, 2939, 2865, 1741, 1715, 1684, 1612, 1513, 1262 cm⁻¹; ¹H NMR (300 MHz, CD₃CN/CDCl₃, 70:30) δ 7.34–6.72 (m, 17 H), 5.70 (bd, 1 H, J = 6.3 Hz), 5.33 (bd, 1 H, J = 7.0 Hz), 5.05 (d, 1 H, J = 12.6 Hz), 4.98 (d, 1 H, J = 12.6 Hz), 4.65–4.58 (m, 1 H), 4.40–4.24 (m, 2 H), 3.75 (s, 3 H), 3.74 (s, 3 H), 3.62 (s, 3 H), 3.08–2.73 (m, 6 H), 1.38 (s, 9 H).

(R)-3-[4-[2-[[(1,1-Dimethylethoxy)carbonyl]amino]-2-[[[[(2-iodoethyl)oxy]carbonyl]methyl]amino]carbonyl]ethyl]phenoxy]-N-[(phenylmethoxy)carbonyl]-D-phenylglycine α -Methyl Ester (9b). To a stirred solution of the corresponding bromide¹ (190 mg, 0.26 mmol) in 10 mL of dry acetone was added 192 mg (5.0 equiv) of NaI (anhydrous 99+%) and the resulting slurry was heated at reflux for 7 h under N_2 . The reaction mixture was cooled to rt and filtered, and the filter cake was washed well with acetone (10 mL \times 3). The combined organic extracts were evaporated to provide a solid residue which was dissolved in 50 mL of CH₂Cl₂, washed with brine, dried over MgSO4, and concentrated in vacuo. The crude product was flashchromatographed on silica gel (Hex/EtOAc, 6/4) to give 166 mg (82%) of 9b as a white solid: mp 46.0-48.0 °C; $[\alpha]^{26}$ -1.8° (c 0.56, CHCl₃); R_f 0.28 (Hex/EtOAc, 6/4); IR (CHCl₃) 3430, 3019, 1734, 1718, 1685, 1507, 1220 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.34-6.89 (m, 13 H), 6.67-6.65 (m, 1 H, CONHCH₂CO₂), 5.97 (bs, 1 H, NHCO₂CH₂Ph), 5.36 (d, 1 H, J = 6.1 Hz, CHNHCO₂- CH_2Ph), 5.19 (d, 1 H, J = 7.7 Hz, $NHCO_2(CCH_8)_8$), 5.12 (d, 1 H, J = 12.4 Hz, PhCHHOCONH), 5.07 (d, 1H, J = 12.4 Hz, PhCHHOCONH), 4.42-4.35 (m, 3 H, CHNHCO₂C(CH₃)₃ overlapping with CO₂CH₂CH₂I), 4.02-3.96 (m, 2 H, NHCH₂CO₂), 3.72 (s, 3 H, ArCHCO₂CH₃), 3.28-3.24 (m, 2 H, CO₂CO₂CH₂I), 3.14-2.98 (m, 2 H, ArCH₂CHCONH), 1.40 (s, 9 H, NHCO₂C(CH₃)₃); ¹³C NMR (75 MHz, CDCl₃) δ 171.6, 170.9, 168.8, 157.7, 155.4, 155.3, 138.4, 136.0, 131.8, 130.7, 130.1, 128.4, 128.1, 121.6, 119.2, 118.2, 117.3, 80.2, 67.1, 65.2, 57.5, 55.6, 52.9, 41.1, 37.6, 28.2, -0.5.Anal. Calcd for C35H40N3O10I: C, 53.22; H, 5.11. Found: C, 52.82; H, 5.24.

(*R*)-3-[4-[2-[[(1,1-Dimethylethoxy)carbonyl]amino]-2-[[(carboxymethyl)amino]carbonyl]ethyl]phenoxy]-*N*-[(phenylmethoxy)carbonyl]-D-phenylglycine α -Methyl Ester (9c): yield 38%; [α]²³_{Hg,436} +2.4° (c 0.34, MeOH); IR (CHCl₈) 3426, 3020, 2961, 2927, 1724, 1675, 1507, 1217 cm⁻¹; ¹H NMR (300 MHz, CD₃CN + CDCl₃) δ 7.42-6.78 (m, 13 H), 6.37-6.32 (m, 1 H, CONHCH₂CO₂), 5.42-5.26 (m, 2 H, NHCO₂CH₂Ph overlapping with CHNHCO₂CH₂Ph), 5.11 (d, 1 H, J = 12.6 Hz, NHCO₂CHHPh), 5.03 (d, 1 H, J = 12.6 Hz, NHCO₂CHHPh), 4.38-4.28 (m, 2 H, NHBoc overlapping with CHNHBoc), 3.89-3.75 (m, 2 H, NHCH₂CO₂), 3.69 (s, 3 H, CO₂CH₃), 3.10-2.86 (m, 2 H, ArCH₂), 1.38 (s, 9 H, Boc).

3-(4-Chlorophenyl)propionic acid 2-Iodoethyl Ester (10b). Prepared as described for compound 1b: yield 80%; R_f 0.49 (Hex/ EtOAc, 8/2); IR (CHCl₃) 3019, 1735, 1493 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.26 (d, 2 H, J = 8.3 Hz), 7.14 (d, 2 H, J = 8.3 Hz), 4.32 (t, 2 H, J = 6.8 Hz), 3.26 (t, 2 H, J = 6.8 Hz), 2.94 (t, 2 H, J = 7.6 Hz), 2.65 (t, 2 H, J = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 138.6, 132.0, 129.6, 128.5, 64.4, 35.3, 30.0, 0.4; HRMS calcd for C₁₁H₁₂O₂CII 337.9572, found 337.9573.

3-(4-Chlorophenyl)propionic acid (10c): yield 90%; mp 120.5-122.0 °C (lit.¹⁰ 122-123 °C); R_f 0.45 (MeOH/CHCl₃, 1/9); IR (CHCl₃) 3019, 2949, 1712, 1493, 1409 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) d 7.27 (d, 2 H, J = 8.5 Hz), 7.14 (d, 2 H, J = 8.5 Hz), 2.93 (t, 2 H, J = 7.4 Hz), 2.67 (t, 2 H, J = 7.4 Hz).

n-Octanoic Acid 2-Iodoethyl Ester (11b). Prepared as described for compound 1b: yield 75%; R_f 0.47 (Hex/CH₂Cl₂, 1/1); IR (CHCl₃) 2955, 2930, 2857, 1734, 1602, 1468, 1220 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.33 (t, 2 H, J = 6.8 Hz), 3.30 (t, 2 H, J = 6.8 Hz), 2.34 (t, 2 H, J = 7.5 Hz), 1.66–1.62 (m, 2 H), 1.30–1.28 (m, 8 H), 0.88 (t, 3 H, J = 6.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 173.1, 64.2, 34.1, 31.6, 29.0, 28.8, 24.8, 22.5, 14.0, 0.5; HRMS calcd for C₁₀H₁₉O₂I 298.0432, found 298.0434.

n-Octanoic acid (11c): yield 93%; IR (CHCl₃) 2958, 2930, 1708 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 2.35 (t, 2 H, J = 7.5 Hz), 1.66–1.59 (m, 2 H), 1.31–1.28 (m, 8H), 0.88 (t, 3 H, J = 6.8 Hz).

3-(Benzyloxy)phenylacetic Acid 2-Iodoethyl Ester (12b). Prepared as described for compound 1b: yield 73%; R_f 0.49 (Hex/ EtOAc, 8/2); IR (CHCl₃) 3019, 1737, 1600, 1519 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.22 (m, 5 H), 6.93–6.88 (m, 3 H), 5.06 (s, 2 H), 4.33 (t, 2 H, J = 6.9 Hz), 3.62 (s, 2 H), 3.26 (t, 2 H, J= 6.9 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 170.6, 158.8, 136.8, 134.9, 129.5, 128.5, 127.9, 127.4, 121.8, 115.8, 113.6, 69.8, 64.8, 41.1, 0.2; HRMS calcd for C₁₇H₁₇O₃I 396.0224, found 396.0226.

3-(Benzyloxy)phenylacetic acid (12c): yield 88%; mp 123.0–125.0 °C; R_f 0.52 (EtOAc/MeOH, 9/1); IR (CHCl₃) 3012, 2917, 1712, 1598 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.42–7.18 (m, 5 H), 6.89–6.84 (m, 3 H), 5.02 (s, 2 H), 3.60 (s, 2 H); ¹³C NMR (75 MHz, CHCl₃) δ 177.6, 159.0, 136.8, 134.6, 129.7, 128.6, 128.0, 127.5, 122.0, 116.0, 113.7, 90.0, 41.0; HRMS calcd for C₁₅H₁₄O₃ 242.0943, found 242.0941.

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Supplementary Material Available: Copies of ¹H and ¹³C NMR spectra of all compounds (54 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

⁽⁸⁾ Pearson, A. J.; Lee, K. J. Org. Chem., submitted.
(9) Evans, D. A.; Ellman, J. A. J. Am. Chem. Soc. 1989, 111, 1063.

⁽¹⁰⁾ Wittstruck, T. A.; Trachtenberg, G. N. J. Am. Chem. Soc. 1969, 89, 3803.